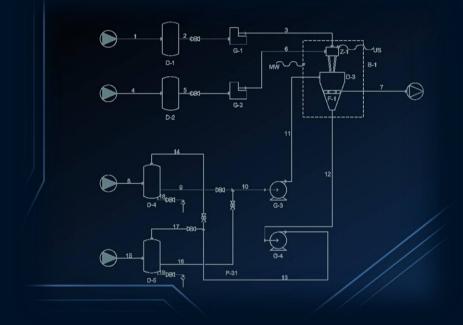
Novel technologies and process intensification in the production of micro-systems with pharmacological/nutraceutical activity



Annalisa Dalmoro

Novel technologies and process intensification in the production of micro-systems with pharmacological/nutraceutical activity

Annalisa Dalmoro







FONDO SOCIALE EUROPEO

Programma Operativo Nazionale 2007/2013 "Ricerca Scientifica, Sviluppo Tecnologico, Alta Formazione" Regioni dell'Obiettivo 1 – Misura III.4 "Formazione superiore ed universitaria"

Department of Industrial Engineering

Corso di dottorato in Scienza e tecnologie per l'industria chimica, farmaceutica e alimentare curriculum Ingegneria Chimica (XI ciclo)

Novel technologies and process intensification in the production of micro-systems with pharmacological/nutraceutical activity

Supervisors Prof. Matteo d'Amore Prof. Anna Angela Barba **Ph.D. student** Annalisa Dalmoro

Scientific Referees

Prof. Roland Bodmeier (Freie Universität Berlin) Prof. Nadia Passerini (Università di Bologna)

Ph.D. Course Coordinator *Prof. Paolo Ciambelli*

Una ricerca comincia sempre con la Fortuna del Principiante. E finisce sempre con la Prova del Conquistatore.

Paulo Coelho, L'alchimista, 1988

Publications

(Inherent the Ph.D. project)

- <u>Dalmoro A.</u>; Barba A.A.; d'Amore M.; Lamberti G.; (2012), "Microparticles production by a single-pot semi-continuous bench scale apparatus", submitted to AIChE Journal;
- Barba A.A.; **Dalmoro A.**; d'Amore M.; Lamberti G.; "Controlled release of drugs from microparticles produced by ultrasonic assisted atomization based on biocompatible polymers", Chem. Biochem. Eng. Q., 26(4) 345-354 (2012);
- Barba A.A.; **Dalmoro A.**; d'Amore M.; (2012) "An engineering approach to biomedical sciences: advanced strategies in drug delivery systems production", Translational Medicine @ UniSa, 4 (1) 5-11 (2012);
- <u>Dalmoro A.</u>; Barba A.A.; Lamberti G.; Grassi M.; d'Amore M.; "Pharmaceutical applications of biocompatible polymer blends containing sodium alginate", Advances in Polymer Technology, 31(3) 219-230 (2012);
- **Dalmoro A.**; Barba A.A.; Lamberti G.; d'Amore M.; "Intensifying the microencapsulation process: Ultrasonic atomization as an innovative approach", European Journal of Pharmaceutics and Biopharmaceutics, 80 471–477 (2012);
- <u>Dalmoro A.</u>; Barba A.A.; d'Amore M.; Lamberti G.; "Micro-Systems Production: A Promising New Technique with Low Energy Consumption", Scientia Pharmaceutica, 78 (3) 670-670 (2010);

Proceedings

- <u>A. Dalmoro</u>, A.A. Barba, G. Lamberti, M. d'Amore (2012), Sistemi particellari shell-core prodotti via atomizzazione assistita da ultrasuoni, *Convegno GRICU 2012* Montesilvano (PE) ITALY, 16-19 settembre 2012
- S. Cascone, <u>A. Dalmoro</u>, G. Lamberti, A.A. Barba (2012), Metodi innovativi di preparazione e testing per sistemi farmaceutici, *Convegno GRICU 2012* Montesilvano (PE) ITALY, 16-19 settembre 2012
- <u>A. Dalmoro</u>, M. d'Amore, A.A. Barba, Shell-core particles production by coaxial double channel device, presented to 8th World Meeting on Pharmaceutics, Biopharmaceutics and Pharmaceutical Technology, 19th-22nd March 2012, Istanbul (Turkey)
- I. Galdi, <u>A. Dalmoro</u>, G. Lamberti, G. Titomanlio, A.A. Barba, M. d'Amore, Modeling of the controlled drug release from solid matrices based on swellable/erodible polymeric hydrogels, presented to 19th International Congress of Chemical and Process Engineering CHISA 2010 and the 7th European Congress of Chemical Engineering ECCE-7, 28th August 1st September 2010, Prague (Czech Republic)
- <u>A. Dalmoro</u>, I. Galdi, G. Lamberti, G. Titomanlio, A.A. Barba, M. d'Amore, Targeted oral drug delivery by pH-sensitive microparticles, presented to 19th International Congress of Chemical and Process Engineering CHISA 2010 and the 7th European Congress of Chemical Engineering ECCE-7, 28th August 1st September 2010, Prague (Czech Republic)
- I. Galdi, <u>A. Dalmoro</u>, G. Lamberti, G. Titomanlio, A.A. Barba, M. d'Amore, Swelling, erosion and drug release in hydrogel based solid matrices, presented to 7th World Meeting on Pharmaceutics, Biopharmaceutics and Pharmaceutical Technology, 8th-11th March 2010, Valletta (Malta).
- Dalmoro, I. Galdi, G. Lamberti, G. Titomanlio, A.A. Barba, M. d'Amore. "PH-sensitive microparticles for enteric drug delivery by solvent evaporation from double emulsion", presented to 7th World Meeting on Pharmaceutics, Biopharmaceutics and Pharmaceutical Technology, 8th-11th March 2010, Valletta (Malta).

Contents

Publications	iii
Proceedings	iii
Contents	I
Figures index	V
Tables index	IX
Abstract	XI
Introduction	1
1.1 Microencapsulation	2
1.1.1 Importance of microencapsulation and applications	2
1.1.2 Fundamental steps of microencapsulation	3
1.1.3 Scale of production	6
1.2 Process intensification	9
1.1 Aims of the PhD thesis	10
1.2 Outline of the thesis	11
State of the art	13
2.1 Microencapsulation apparatuses	14

2.	1.1 Patents					_ 14
2.	1.2 Other literature microencapsulat		-		ses for	_ 28
	Apparatuses					
	mization)					
2.	2.1 Principles of th	e ult	rasonic	atomizatio	n and benefits	_ 31
	2.2 Spray-drying u	Ũ				
2.	2.3 Coaxial ultraso	nic a	tomize	r		_ 37
2.3	Remarks about	: sta	te of t	he art	•••••	41
Ult	rasonic atomiza	tion	phen	omena	•••••	43
3.1	Atomization	•••••	•••••	•••••	••••••	44
3.	1.1 Dripping					_ 46
3.	1.2 Ultrasonic aton	nizati	ion			_ 47
Mie	croencapsulatio	n ap	oparat	tus buildi	ng	51
4.1	General descrij	otio	n of th	ne appara	atus	52
4.	1.1 The apparatus:	first	stage_			_ 52
4.	1.2 The apparatus:	final	state_			_ 54
4.2	Criteria of com	pon	ents s	election.	•••••	56
4.	2.1 Atomizing sect	ion _				_ 56
4.	2.2 Feeding section	1				_ 58
4.	2.3 Separation/dryi	ng se	ection _			_ 59
4.3	Description of J	plan	t opei	ration	••••••	60
Mie	croencapsulatio	n ap	oparat	tus testin	g	63
5.1	Process parame	eters	s defiı	nition	••••••	64
5.	1.1 Operative para	nete	rs: feed	ing		_ 64

5.1.1.a Materials: alginate	64
5.1.1.b Selection of feeding parameters	67
5.1.2 Operative parameters: atomization	71
5.1.3 Operative parameters: separation and stabilization	71
5.2 Encapsulation of model molecules	72
5.3 Materials	73
5.3.1 Vitamin B12 selection	73
5.3.2 α-tocopherol selection	
5.4 Methods	75
5.4.1. Vitamin B12 particles manufacture	75
5.4.2 Vitamin B12 loading and release tests	77
5.4.3 α-tocopherol particles manufacture	77
5.4.4 α-tocopherol particles characterization	78
5.4.4.a Image analysis	78
5.4.4.b Dielectric properties	78
5.4.4.c Moisture and temperature measurements	79
5.4.4.d Differential scanning calorimetry	79
5.4.4.e α -tocopherol loading and release tests	79
5.4.4.f Apparatus power consuming	80
5.5 Preliminary results (vitamin B12 encapsulation	n)81
5.5.1 Steps towards atomization	87
5.6 Results about final encapsulating tests	92
5.6.1 Droplets size prediction by literature correlations	101
5.6.1.a Dripping	102
5.6.1.b Ultrasonic atomization	103
Concluding remarks	107

6.1 Concluding remarks	108
Appendix A	113
A.1 Assay methods: UV-vis spectrometry vs HPL0	C 114
A.1.1 Materials and Methods	_ 114
A.1.2 UV-vis spectrometer: method of spectra subtraction_	_ 117
A.1.3 HPLC analysis	_ 120
A.1.4 Choice of the most reliable method	_ 122
Appendix B	125
B.1 Diffusion in a sphere	126
References	129
CURRICULUM VITAE	137

Figures index

Figure 1. Process intensification toolbox [31]	9
Figure 2. Idealization of the intensified plant of microencapsulation	. 11
Figure 3. Apparatus using a hydraulic piston for obtaining a sudden pressure change to achieve microencapsulation [33]	. 14
Figure 4. Ultrasonic apparatuses for microencapsulation [33]	. 15
Figure 5. Scheme of the plant for the production of alginate pellets [34]	. 17
Figure 6. Two variations of the process for microspheres preparation [35]	. 18
Figure 7. Equipment configuration for preparing micro-particles using a static mixing assembly [8]	. 20
Figure 8. Scheme of the longitudinal cross-section of a mixing block [36]	. 20
Figure 9. Global scheme of a system comprising a mixing-block [36]	. 21
Figure 10. Different configurations of liquid-liquid extraction apparatus [26]	. 22
Figure 11. Process flow diagram of a system for micro-particles production using a contact tank with recycle, in a continuous process [38]	. 23
Figure 12. Scheme of the apparatus for producing micro-particles containing nucleic-acid [39]	. 25
Figure 13. Scheme of three different apparatuses to produce micro-particles, including a freezing zone [38]	. 26
Figure 14. Flow diagram of the continuous spray-capture microencapsulation process [40]	. 28
Figure 15. BRACE microencapsulation process [41]	. 29
Figure 16. Apparatus for co-axial electrohydrodynamic atomization [42]	. 30
Figure 17. Ultrasonic frequency ranges, kind of applications and ultrasonic atomization mechanism sketch [45]	. 31
Figure 18. Scheme of an ultrasonic atomizer [53]	. 35
Figure 19. Sketch of the vacuum spray-dryer (A) with detailed view of the ultrasonic nozzle (B)	. 36

Figure 20. Scheme of the ultrasound/spray-drying system [55]
Figure 21. Schematic description of the microencapsulation system using a coaxial ultrasonic atomizer [58]
Figure 22. Precision particle fabrication apparatus to produce uniform double- walled microspheres [57]
Figure 23. The three sections of pumping/feeding, atomization and stabilization 52
Figure 24. A scheme of the initial state of the apparatus
Figure 25. Scheme of the single-pot semi-continuous bench scale apparatus
Figure 26. Dimensional features, in mm, of wet-collector, with the details of both fluid dispenser (B) and filter (C)
Figure 27. Cross-sectional view of a typical nozzle [66]57
Figure 28. Dual liquid feed assembly [66]58
Figure 29. Apparatus parameters to be controlled and optimized64
Figure 30. Structure of alginate: chemical formula of MM, GM, GG blocks [72] 65
Figure 31. Molecular models of two GG dimers (up) and of two MM dimers (down) with Ca ²⁺ [74]
Figure 32. Coaxial dripping to obtain shell-core systems
Figure 33. Example of shell-core particles: blue core confined in the transparent shell material70
Figure 34. Shell-core particle and only-core (or matrix) particle72
Figure 35. Vitamin B12 structure [81]74
Figure 36. Structure of α-tocopherol [81]75
Figure 37. Parameters for particles manufacture by dripping76
Figure 38. Pictures of fresh particles, both shell-core (up) and only-core (down), and of dried particles (CD: convective; MW: microwave)
Figure 39. Fraction of B12 released in a phosphate buffer at pH 7.4, particles obtained by dripping, without drying (fresh particles)
Figure 40. Fraction of B12 released in a phosphate buffer at pH 7.4, particles obtained by first dripping and then drying in a tray-oven with air at 45°C
Figure 41. Fraction of B12 released in a phosphate buffer at pH 7.4, particles obtained by first dripping and then drying in a microwave cavity
Figure 42. Mass of B12 released in the phosphate buffer at pH 7.4 for fresh, convective dried, and microwave dried particles
Figure 43. Fraction of B12 released in two step test, particles obtained by dripping without drying (fresh particles)

Figure 44. Fraction of B12 released in two step test, particles obtained by first dripping and then drying in a tray-oven with air at 45°C
Figure 45. Fraction of B12 released in two step test, particles obtained by first dripping and then drying in a microwave cavity
Figure 46. Optical microscope picture of shell-core micro-particles obtained by coaxial ultrasonic atomization (P=4W)
Figure 47. Drop diameter distribution for 25 KHz ultrasonic atomizer (data for water) [66]
Figure 48. HPLC signals deriving from HPLC analysis of supernatant and washing water of both shell-core and matrix preparations
Figure 49. HPLC signals of pH 1 solutions (first part of two step release test) after 10 minutes of dissolution of respectively shell-core (dotted line) and only core (solid line) micro-particles
Figure 50. Optical microscope pictures of shell-core micro-particles: fresh (down) and dried (up)
Figure 51. Comparison between dielectric properties of water (triangles) and those of produced micro-particles (circles) at room temperature
Figure 52. UP: DSC scans of pure alginate, cross-linking powder CaCl ₂ , physical mix of alginate and CaCl ₂ ; DOWN: DSC scans of pure alginate, particles of cross-linked alginate obtained by atomization and then dried by convective drying (CD) or by microwave drying (MW)
Figure 53. α-tocopherol released (amount of TOC released and measured/theoretical loaded amount of TOC) from alginate matrix and shell-core micro-particles, produced by ultrasonic atomization and stabilized by convective drying (up) or by microwave drying (down)
Figure 54. α-tocopherol released (amount of TOC released and measured/theoretical loaded amount of TOC) from alginate matrix and shell-core macro-particles (beads), stabilized by convective drying (up) or by microwave drying (down)
Figure 55. α-tocopherol released (amount of TOC released and measured/theoretical loaded amount of TOC) from alginate matrix micro (stars) and macro-particles (squares), stabilized by convective drying (up) or by microwave drying (down)
Figure 56. Repartition of power consumed among atomization (2.9%), hardening and separation (1.6%), drying (95.4%). This percentage is defined as the ratio between the specific energies, (joule)/(g of fresh product), of a single section (for example, atomization section) and the sum of the three sections 100
Figure 57. Nozzle geometry and relevant dimensions
Figure 58. Spectra of alginate particles, dried by convective drying (CD) or by microwave drying (MW), and put in three solutions (pH1, bicarbonate buffer

and phosphate buffer, both at pH 7.4) at two different concentrations (0.5 g/l; 1 g/l)	117
Figure 59. Absorbance spectrum (solid line), exponential (dashed line), and curve obtained by subtraction (dotted line), for a system alginate-B12 obtained by dripping it in the cross-linking solution and drying in a tray oven with air at 45°C, and then dissolved in a phosphate buffer at pH 7.4	118
Figure 60. Absorbance spectrum (solid line), exponential (dashed line), and curve obtained by subtraction (dotted line), for a system alginate-B12 obtained by atomizing it in the cross-linking solution, drying in a tray-oven with air at 45°C, and then dissolved in a phosphate buffer at pH 7.4. Top: full spectrum; down: particular of spectrum in the range 300-400 nm	119
Figure 61. Signals given by HPLC analysis, using the three different separation procedures	121

Tables index

Table 1. Example of some microencapsulated molecules [1]
Table 2. Main methods for micro-particles preparation [3]4
Table 3. A comparison between the features of static and CSTR mixers [20]
Table 4. Microencapsulation processes and their applicabilities [1]7
Table 5. PLGA based micro-particle formulations available on the market [29] 8
Table 6. Variable flow rates, mL/min, according to the combination of drive options and tube diameters [67]
Table 7. Typical properties of alginate Manugel GHB [79] 68
Table 8. Polymer (alginate) concentrations (w/w) of both core and shell solutions and their effect
Table 9. Parameters selected for α-tocopherol particles production
Table 10. Size mean and standard deviation for both shell-core and matrix particles, fresh and dried (CD or MW); shrinkage percentage for the dried ones 81
Table 11. Mean size and standard deviation for both shell-core and matrix micro-particles, fresh and dried; volumetric shrinkage percentage for the dried ones
Table 12. Physical properties of Manugel GHB alginate solutions [79]: the properties of a solution with a concentration of 1.5% (w/w) are highlighted 101
Table 13. Values to insert in correlations for droplet size prediction in ultrasonic atomization 103
Table 14. Features of the three different HPLC procedures 121
Table 15. Comparison among different ways for detection of vitamin B12 in asystem containing alginate (particles obtained by dripping)122
Table 16. Comparison among different ways for detection of vitamin B12 in a system containing alginate (particles obtained by atomization)

Abstract

Purpose of the PhD thesis was to develop a novel microencapsulation process, designing and building a single-pot semi-continuous bench scale apparatus. The novel process is based on the coupling of two emerging techniques, involving ultrasound and microwave, used in atomization and heating operations, respectively. The process has been designed to respond to the needs for process intensification, i.e. improvement of process efficiency and cutting down of energy consumption. With this aim, a review of the main processes used for microencapsulation was first performed: conventional processes showed a number of drawbacks, such as high energy consumption, batch configuration, use of solvents and long times of production. On the basis of the state of the art, the idea of an intensified apparatus for particles production, exploiting alternative resources, such as ultrasound and microwave, was formulated. The apparatus was composed of three main sections: feeding, atomization. separation/stabilization. The feeding and atomization sections were built connecting a double channel ultrasonic atomizer to a system for feeding solutions in a purposely designed separation/stabilization section, thus realizing a semi-continuous apparatus. Separation section consisted of a wet-collector, i.e. a sort of hydrocyclone, which allowed a uniform distribution of the hardening solution and the consequent contact with the atomized drops, a filtering device, and a microwave oven. The wet-collector was placed into the microwave oven to obtain an "on-line" drying. Recirculation of the hardening solution, to renew contact surface between droplets and cross-linker, was guaranteed by system of centrifugal pumps. In this configuration, when a atomization occurred, drops were harvested in the wet-collector. After atomization, the obtained suspension was collected in the cross-linker tank, then the filtering device was inserted in the lower part of the wet-collector, so that hardening solution was recovered and particles

settled on the filter, when the suspension was brought again to the wet-collector and after its complete emptying. An eventual following washing step can be done in a similar way to the previous hardening step. Finally, particles were stabilized by microwave drying, and then recovered.

The steps for building the microencapsulation apparatus were accurately shown. Then, criteria used for components selection, in order to obtain the best performances from the plant, were highlighted. After building the plant, the process parameters were defined. First, the research for the best combination of feeding parameters, such as type of materials, composition, concentration and feed rate, that assure the encapsulation of the core material in the shell, was carried out. Then, the parameters of the ultrasonic atomizer (atomization section), essentially power, were tuned. Finally, for stabilization/separation section, fundamental was the relevant stabilization step, where microwave power was set to avoid too high temperatures that could degrade molecules.

The ability of the novel plant to obtain micronized systems, that exhibit a behavior interesting for the pharmaceutical or nutraceutical markets, was tested. Micro-particles characterization showed that it is possible to obtain a shell-core configuration encapsulating two functional molecules, vitamin B12 and α -tocopherol. Some important results were: 1) high loading and enteric (gastro-resistant) behavior of micro-particles; 2) delayed release for shell-core micro-systems compared to matrix ones; shell-core configuration in macro-scale (beads) able to prevent degradation of α -tocopherol, instead observed in matrix beads. Moreover, microwave treatment (not harsher for the short irradiation times) caused, especially for shell-core configuration, a little delay in molecule release. Resuming, better release properties for systems produced in the novel apparatus were achieved by the of atomization drying. coupling ultrasonic and microwave Furthermore, the basic transport phenomena occurring in the ultrasonic assisted atomization were investigated, emphasizing the role of operative parameters, and literature correlations, based on forces balance, were also applied for droplets size prediction. All these results endorses the usefulness of the novel plant, based on the combination of two powerful tools of process intensification, ultrasonic atomization and microwave drying, to obtain microparticularly systems. interesting for specific drug delivery

applications. Moreover, working at room conditions and in absence of solvents, improving the energy transfer rate (faster process times), reducing process chambers volume (low particles inertia in ultrasonic atomization, single-pot process realization), enhancing the product quality (micro-particles with tailored features), makes the apparatus more attractive in terms of improved inherent safety and reduced costs.

References

[1] P. Venkatesan, R. Manavalan, K. Valliappan, Microencapsulation: A vital technique in novel drug delivery system, Journal of Pharmaceutical Sciences, 1 (2009) 26-35.

[2] J. Silva, Effect of drug properties on the release from CAP microspheres prepared by a solvent evaporation method, Journal of microencapsulation, 16 (1999) 95-103.

[3] P. Colombo, P. Catellani, A. Gazzaniga, E. Menegatti, E. Vidale, Principi di tecnologie farmaceutiche, Casa Editrice Ambrosiana, Milano, (2004).

[4] M. Ré, B. Biscans, Preparation of microspheres of ketoprofen with acrylic polymers by a quasi-emulsion solvent diffusion method, Powder Technology, 101 (1999) 120-133.

[5] N. Barakat, A. Ahmad, Diclofenac sodium loaded-cellulose acetate butyrate: Effect of processing variables on microparticles properties, drug release kinetics and ulcerogenic activity, Journal of microencapsulation, 25 (2008) 31-45.

[6] S. Benita, Microencapsulation: methods and industrial applications, Informa HealthCare, 2006.

[7] S. Freitas, H. Merkle, B. Gander, Microencapsulation by solvent extraction/evaporation: reviewing the state of the art of microsphere preparation process technology, Journal of controlled Release, 102 (2005) 313-332.

[8] S. Lyons, S. Wright, Apparatus and method for preparing microparticles, in, 2003.

[9] J. Herrmann, R. Bodmeier, Biodegradable, somatostatin acetate containing microspheres prepared by various aqueous and non-aqueous solvent evaporation methods, European Journal of Pharmaceutics and Biopharmaceutics, 45 (1998) 75-82.

[10] J. Beyger, J. Nairn, Some factors affecting the microencapsulation of pharmaceuticals with cellulose acetate phthalate, Journal of Pharmaceutical Sciences, 75 (1986) 573-578.

[11] F. Gabor, Ketoprofen-poly (D, L-lactic-co-glycolic acid) microspheres: influence of manufacturing parameters and type of

polymer on the release characteristics, Journal of microencapsulation, 16 (1999) 1-12.

[12] Y. Yang, T. Chung, N. Ping Ng, Morphology, drug distribution, and in vitro release profiles of biodegradable polymeric microspheres containing protein fabricated by double-emulsion solvent extraction/evaporation method, Biomaterials, 22 (2001) 231-241.

[13] W. Obeidat, J. Price, Preparation and evaluation of Eudragit S 100 microspheres as pH-sensitive release preparations for piroxicam and theophylline using the emulsion-solvent evaporation method, Journal of microencapsulation, 23 (2006) 195-202.

[14] H. Chen, J. Wu, H. Chen, Preparation of ethylcellulose microcapsules containing theophylline by using emulsion non-solvent addition method, Journal of microencapsulation, 12 (1995) 137-147.

[15] V.S. Mastiholimath, P.M. Dandagi, S.S. Jain, A.P. Gadad, A.R. Kulkarni, Time and pH dependent colon specific, pulsatile delivery of theophylline for nocturnal asthma, International Journal of Pharmaceutics, 328 (2007) 49-56.

[16] I. Péter Sipos, Influence of preparation conditions on the properties of Eudragit microspheres produced by a double emulsion method, Drug Development Research, 64 (2005) 41-54.

[17] M. Lai, R. Tsiang, Microencapsulation of acetaminophen into poly (L-lactide) by three different emulsion solvent-evaporation methods, Journal of microencapsulation, 22 (2005) 261-274.

[18] P. Sansdrap, A. Mo s, Influence of manufacturing parameters on the size characteristics and the release profiles of nifedipine from poly (DL-lactide-co-glycolide) microspheres, International Journal of Pharmaceutics, 98 (1993) 157-164.

[19] Y. Maa, C. Hsu, Microencapsulation reactor scale-up by dimensional analysis, Journal of microencapsulation, 13 (1996) 53-66. [20] Y. Maa, C. Hsu, Liquid-liquid emulsification by static mixers for use in microencapsulation, Journal of microencapsulation, 13 (1996) 419-433.

[21] B. Amsden, The production of uniformly sized polymer microspheres, Pharmaceutical research, 16 (1999) 1140-1143.

[22] H. Liu, Science and engineering of droplets: fundamentals and applications, William Andrew, 2000.

[23] M. Rawat, S. Saraf, Influence of selected formulation variables on the preparation of enzyme-entrapped eudragit S100 microspheres, AAPS PharmSciTech, 8 (2007) 289-297.

[24] M. Kitajima, T. Yamaguchi, A. Kondo, N. Muroya, Encapsulation method, in, 1972.

[25] M. Li, O. Rouaud, D. Poncelet, Microencapsulation by solvent evaporation: State of the art for process engineering approaches, International Journal of Pharmaceutics, 363 (2008) 26-39.

[26] J. Ramstack, Apparatus and method for preparing microparticles using liquid-liquid extraction, in, 2002.

[27] M. Rickey, J. Ramstack, D. Lewis, Preparation of biodegradable, biocompatible microparticles containing a biologically active agent, in, 2003.

[28] S. Gouin, Microencapsulation: industrial appraisal of existing technologies and trends, Trends in Food Science & Technology, 15 (2004) 330-347.

[29] C. Wischke, S. Schwendeman, Principles of encapsulating hydrophobic drugs in PLA/PLGA microparticles, International Journal of Pharmaceutics, 364 (2008) 298-327.

[30] J. Coulson, J. Richardson, Chemical Engineering, Volume 2: Particle Technology and Separation Processes, Pergamon, (1991).

[31] A. Stankiewicz, J. Moulijn, Re-engineering the chemical processing plant: process intensification, M. Dekker, New York, 2004. [32] G. Cravotto, A. Barge, L. Boffa, E. Calcio Gaudino, W. Bonrath, Ultrasound-microwave coupling: an efficient tool for chemical-process intensification, in, GPE-EPIC, 2nd International Congress on Green Process Engineering, 2nd European Process Intensification Conference, 2009.

[33] B. Redding Jr, Apparatus and method for making microcapsules, in, 1993.

[34] G. Alisch, E. Brauneis, B. Pirstadt, N. Iffland, E. Brandau, Process and plant for the production of spherical alginate pellets, in, 1995.

[35] B. Amsden, R. Liggins, Methods for microsphere production, in, 2001.

[36] D. Bomberger, P. Catz, M. Smedley, P. Stearns, System and method for producing drug-loaded microparticles, in, 2002.

[37] J. Gibson, R. Holl, A. Tipton, Emulsion-based processes for making microparticles, in, 2002.

[38] P. Herbert, M. Healy, Production scale method of forming microparticles, in, 2004.

[39] M. Tyo, Y. Hsu, M. Hedley, Continuous-flow method for preparing microparticles, in, 2004.

[40] J. Piechocki, D. Kyle, Continuous spray-capture production system, in, 2009.

[41] T. Brandau, Preparation of monodisperse controlled release microcapsules, International Journal of Pharmaceutics, 242 (2002) 179-184.

[42] R. Pareta, M. Edirisinghe, A novel method for the preparation of biodegradable microspheres for protein drug delivery, Journal of The Royal Society Interface, 3 (2006) 573.

[43] M. Chang, E. Stride, M. Edirisinghe, Controlling the thickness of hollow polymeric microspheres prepared by electrohydrodynamic atomization, Journal of The Royal Society Interface, 7 (2010) S451.

[44] Y. Lee, F. Mei, M. Bai, S. Zhao, D. Chen, Release profile characteristics of biodegradable-polymer-coated drug particles fabricated by dual-capillary electrospray, Journal of controlled Release, (2010).

[45] A. Dalmoro, A.A. Barba, G. Lamberti, M. d'Amore, Intensifying the microencapsulation process: Ultrasonic atomization as an innovative approach, European Journal of Pharmaceutics and Biopharmaceutics, (2012).

[46] A. Patist, D. Bates, Ultrasonic innovations in the food industry: From the laboratory to commercial production, Innovative food science & emerging technologies, 9 (2008) 147-154.

[47] D. Ensminger, F. Stulen, Ultrasonics: data, equations, and their practical uses, CRC Press, 2009.

[48] B. Avvaru, M. Patil, P. Gogate, A. Pandit, Ultrasonic atomization: effect of liquid phase properties, Ultrasonics, 44 (2006) 146-158.

[49] R. Rajan, A. Pandit, Correlations to predict droplet size in ultrasonic atomisation, Ultrasonics, 39 (2001) 235-255.

[50] K. Park, Y. Yeo, Microencapsulation using ultrasonic atomizers, in, 2003.

[51] L. Rodriguez, N. Passerini, C. Cavallari, M. Cini, P. Sancin, A. Fini, Description and preliminary evaluation of a new ultrasonic atomizer for spray-congealing processes, International Journal of Pharmaceutics, 183 (1999) 133-143.

[52] N. Ashgriz, Handbook of Atomization and Sprays: Theory and Applications, Springer, 2011.

[53] B. Bittner, T. Kissel, Ultrasonic atomization for spray drying: a versatile technique for the preparation of protein loaded biodegradable microspheres, Journal of microencapsulation, 16 (1999) 325-341.

[54] S. Freitas, H. Merkle, B. Gander, Ultrasonic atomisation into reduced pressure atmosphere--envisaging aseptic spray-drying for microencapsulation, Journal of controlled Release, 95 (2004) 185-195.

[55] P. Luz, A. Pires, O. Serra, A low-cost ultrasonic spray dryer to produce spherical microparticles from polymeric matrices, Química Nova, 30 (2007) 1744-1746.

[56] P. Burke, L. Klumb, J. Herberger, X. Nguyen, R. Harrell, M. Zordich, Poly (lactide-co-glycolide) microsphere formulations of darbepoetin alfa: spray drying is an alternative to encapsulation by spray-freeze drying, Pharmaceutical research, 21 (2004) 500-506.

[57] C. Berkland, E. Pollauf, D. Pack, K. Kim, Uniform double-walled polymer microspheres of controllable shell thickness, Journal of controlled Release, 96 (2004) 101-111.

[58] Y. Yeo, K. Park, A new microencapsulation method using an ultrasonic atomizer based on interfacial solvent exchange, Journal of controlled Release, 100 (2004) 379-388.

[59] R. Graves, D. Poole, R. Moiseyev, L. Bostanian, T. Mandal, Encapsulation of Indomethacin Using Coaxial Ultrasonic Atomization Followed by Solvent Evaporation, Drug development and industrial pharmacy, 34 (2008) 419-426.

[60] J. Legako, N. Dunford, Effect of Spray Nozzle Design on Fish Oil–Whey Protein Microcapsule Properties, Journal of Food Science, 75 (2010) E394-E400.

[61] L. Pilon, H. Berberoglu, Method and apparatus for liquid microencapsulation with polymers using ultrasonic atomization, in, 2005.

[62] H. Liu, Science and Engineering of Droplets:: Fundamentals and Applications, William Andrew, 1999.

[63] R.J. Lang, Ultrasonic atomization of liquids, The journal of the acoustical society of America, 34 (1962) 6-8.

[64] K.A. Ramisetty, A.B. Pandit, P.R. Gogate, Investigations into ultrasound induced atomization, Ultrasonics Sonochemistry, (2012).

[65] A.A. Barba, M. d'Amore, S. Cascone, G. Lamberti, G. Titomanlio, Intensification of biopolymeric microparticles production by ultrasonic assisted atomization, Chemical Engineering and Processing: Process Intensification, 48 (2009) 1477-1483.

[66] <u>www.sono-tek.com</u>, in.

[67] <u>www.verderflex.com</u>, in.

[68] www.watson-marlow.com, in.

[69] R.A. Arterburn, The sizing and selection of hydrocyclones, Design and Installation of Comminution Circuits, (1982) 592-607.

[70] W. Yu, H. Song, G. Zheng, X. Liu, Y. Zhang, X. Ma, Study on membrane characteristics of alginate-chitosan microcapsule with cell growth, Journal of Membrane Science, 377 (2011) 214-220.

[71] M. George, T.E. Abraham, Polyionic hydrocolloids for the intestinal delivery of protein drugs: alginate and chitosan--a review, Journal of Controlled Release, 114 (2006) 1-14.

[72] H. Zhang, H. Wang, J. Wang, R. Guo, Q. Zhang, The effect of ionic strength on the viscosity of sodium alginate solution, Polymers for Advanced Technologies, 12 (2001) 740-745.

[73] W. Gombotz, S. Wee, Protein release from alginate matrices, Advanced Drug Delivery Reviews, 31 (1998) 267-285.

[74] C. DeRamos, A. Irwin, J. Nauss, B. Stout, 13C NMR and molecular modeling studies of alginic acid binding with alkaline earth and lanthanide metal ions, Inorganica chimica acta, 256 (1997) 69-75.

[75] H. Tønnesen, J. Karlsen, Alginate in drug delivery systems, Drug development and industrial pharmacy, 28 (2002) 621-630.

[76] G. Fundueanu, C. Nastruzzi, A. Carpov, J. Desbrieres, M. Rinaudo, Physico-chemical characterization of Ca-alginate microparticles produced with di! erent methods, Biomaterials, 20 (1999) 1435.

[77] M.S. Shoichet, R.H. Li, M.L. White, S.R. Winn, Stability of hydrogels used in cell encapsulation: An in vitro comparison of alginate and agarose, Biotechnology and bioengineering, 50 (1996) 374-381.

[78] A. Dalmoro, A.A. Barba, G. Lamberti, M. Grassi, M. d'Amore, Pharmaceutical applications of biocompatible polymer blends containing sodium alginate, Advances in Polymer Technology, (2012).

[79] E.S. Chan, B.B. Lee, P. Ravindra, D. Poncelet, Prediction models for shape and size of ca-alginate macrobeads produced through extrusion-dripping method, Journal of colloid and interface science, 338 (2009) 63-72.

[80] E. Pasut, R. Toffanin, D. Voinovich, C. Pedersini, E. Murano, M. Grassi, Mechanical and diffusive properties of homogeneous alginate gels in form of particles and cylinders, Journal of Biomedical Materials Research Part A, 87 (2008) 808-818.

[81] Sigma-Aldrich, in.

[82] S.P. Stabler, R.H. Allen, Vitamin B12 deficiency as a worldwide problem, Annu. Rev. Nutr., 24 (2004) 299-326.

[83] F. Aranda, A. Coutinho, M. Berberan-Santos, M. Prieto, J. G mez-Fern ndez, Fluorescence study of the location and dynamics of?tocopherol in phospholipid vesicles, Biochimica et Biophysica Acta-Biomembranes, 985 (1989) 26-32. [84] E.F. Bell, History of vitamin E in infant nutrition, The American Journal of Clinical Nutrition, 46 (1987) 183-186.

[85] M. Simonoska Crcarevska, M. Glavas Dodov, K. Goracinova, Chitosan coated Ca-alginate microparticles loaded with budesonide for delivery to the inflamed colonic mucosa, European Journal of Pharmaceutics and Biopharmaceutics, 68 (2008) 565-578.

[86] A.A. Barba, S. Chirico, A. Dalmoro, G. Lamberti, Simultaneous measurement of theophylline and cellulose acetate phthalate in phosphate buffer by uv analysis, Can J Anal Sci Spectros, 53 (2009) 249-253.

[87] T.W. Wong, L.W. Chan, S.B. Kho, P.W. Sia Heng, Design of controlled-release solid dosage forms of alginate and chitosan using microwave, Journal of controlled Release, 84 (2002) 99-114.

[88] R.E. Mardziah, T.W. Wong, Effects of microwave on drugrelease responses of spray-dried alginate microspheres, Drug development and industrial pharmacy, 36 (2010) 1149-1167.

[89] A. Metaxas, R.J. Meredith, Industrial microwave heating, Inst of Engineering & Technology, 1983.

[90] R.E. Mudgett, Microwave properties and heating characteristics of foods, Food Technology, 40 (1986).

[91] C.M. Sabliov, C. Fronczek, C. Astete, M. Khachaturyan, L. Khachatryan, C. Leonardi, Effects of temperature and UV light on degradation of α -tocopherol in free and dissolved form, Journal of the American Oil Chemists' Society, 86 (2009) 895-902.

[92] T. Pongjanyakul, S. Puttipipatkhachorn, Xanthan–alginate composite gel beads: Molecular interaction and in vitro characterization, International Journal of Pharmaceutics, 331 (2007) 61-71.

[93] A.A. Barba, M. d'Amore, S. Cascone, G. Lamberti, G. Titomanlio, Intensification of biopolymeric microparticles production by ultrasonic assisted atomization, Chemical Engineering and Processing: Process Intensification, 48 (2009) 1477-1483.

[94] H.B. Li, F. Chen, Y. Jiang, Determination of vitamin B12 in multivitamin tablets and fermentation medium by high-performance liquid chromatography with fluorescence detection, Journal of Chromatography A, 891 (2000) 243-247.

[95] E.L. SMITH, K. FANTES, S. BALL, J. WALLER, W. EMERY, B12 Vitamins (Cobalamins), (1951).

[96] Y. Xu, C. Zhan, L. Fan, L. Wang, H. Zheng, Preparation of dual crosslinked alginate-chitosan blend gel beads and in vitro controlled

release in oral site-specific drug delivery system, International Journal of Pharmaceutics, 336 (2007) 329-337. [97] J. Crank, The mathematics of diffusion, (1979).

CURRICULUM VITAE

Ing. Annalisa Dalmoro was born in 01/11/1984. She achieved the bachelor degree in Chemical Engineering, cum laude, in 2006, with a thesis entitled "Analysis of transport phenomena in matrices of swellable hydrogels". Then she gained the Master Degree in Chemical Engineering, cum laude, in 2009 with a thesis entitled "Enteric microparticles coated with smart polymers for controlled drug delivery applications".

In the same year she started her Ph.D. study, described in this thesis, in Science and technologies for chemical, pharmaceutical, and food industry - curriculum Chemical Engineering. She produced several papers, of which some were published on international journals; the remaining being communications to international conferences, and papers published on national journals.

Publications:

- 1. Barba A.A.; <u>Dalmoro A.;</u> d'Amore M.; (2012), "Microwave assisted drying of cellulose derivative (HPMC) granular solids", in press on Powder Technology;
- Barba A.A.; <u>Dalmoro A.</u>; d'Amore M.; Lamberti G.; "Controlled release of drugs from microparticles produced by ultrasonic assisted atomization based on biocompatible polymers", Chem. Biochem. Eng. Q., 26(4) 345-354 (2012);
- 3. Barba A.A.; **Dalmoro A.**; d'Amore M.; Lamberti G.; "In vitro dissolution of pH sensitive micro-particles for colon-specific drug delivery", in press on Pharmaceutical Development and Technology;

- Barba A.A.; <u>Dalmoro A.</u>; d'Amore M.; (2012) "An engineering approach to biomedical sciences: advanced strategies in drug delivery systems production", Translational Medicine @ UniSa, 4 (1) 5-11 (2012);
- <u>Dalmoro A.</u>; Barba A.A.; Lamberti G.; Grassi M.; d'Amore M.; "Pharmaceutical applications of biocompatible polymer blends containing sodium alginate", Advances in Polymer Technology, 31(3) 219-230 (2012);
- <u>Dalmoro A.</u>; Barba A.A.; Lamberti G.; d'Amore M.; "Intensifying the microencapsulation process: Ultrasonic atomization as an innovative approach", European Journal of Pharmaceutics and Biopharmaceutics, 80 471–477 (2012);
- <u>Dalmoro A.</u>; Lamberti G.; Titomanlio G.; Barba A.A.; d'Amore M.; "Enteric Micro-Particles for Targeted Oral Drug Delivery", AAPS PharmSciTech, 11(4) 1500-1507 (2010);
- Dalmoro A.; Barba A.A.; d'Amore M.; Lamberti G.; "Micro-Systems Production: A Promising New Technique with Low Energy Consumption", Scientia Pharmaceutica, 78 (3) 670-670 (2010);
- <u>Dalmoro A.</u>; Villano O.; Barba A.A.; Lamberti G.; "Dosare dove serve", NCF-Notiziario Chimico E Farmaceutico, 49 (4) 112-114 (2010);
- Barba, A.A.; <u>Dalmoro, A.</u>; De Santis, F.; Lamberti, G. "Synthesis and characterization of P(MMA-AA) copolymers for targeted oral drug delivery", *Polymer Bulletin*, 62(5) 679-688 (2009)
- Barba, A.A.; Chirico, S.; <u>Dalmoro, A.</u>; Lamberti, G. "Simultaneous measurement of Theophylline and Cellulose Acetate Phthalate in Phosphate Buffer by UV analysis", Canadian Journal of Analytical Sciences & Spectroscopy, 53(6) 249-253 (2009)
- Chirico S., <u>Dalmoro A.</u>, Lamberti G., Russo G., Titomanlio G., "Analysis and modeling of swelling and erosion behavior for pure HPMC tablet", Journal of Controlled Release, 122 (2) 181-188 (2007)

Proceedings:

Dalmoro A.; Barba A.A.; Lamberti G.; d'Amore M., Sistemi particellari shell-core prodotti via atomizzazione assistita da ultrasuoni, presented to Convegno *GRICU 2012-Ingegneria chimica: dalla nanoscala alla macroscala*), 16th-19th September 2012, Montesilvano (PE) (Italy) - *SPEAKER*

Cascone S.; <u>Dalmoro A.</u>; Lamberti G.; Barba A.A., Metodi innovativi di preparazione e testing per sistemi farmaceutici, presented to Convegno *GRICU 2012-Ingegneria chimica: dalla nanoscala alla macroscala*), 16th-19th September 2012, Montesilvano (PE) (Italy) - *SPEAKER*

Dalmoro A.; d'Amore M.; Barba A.A., Shell-core particles production by coaxial double channel device, presented to 8th World Meeting on Pharmaceutics, Biopharmaceutics and Pharmaceutical Technology, 19th-22nd March 2012, Istanbul (Turkey) - SPEAKER

Galdi I.; <u>Dalmoro A.</u>; Lamberti G.; Titomanlio G.; Barba A.A.; d'Amore M.. "Modeling of the controlled drug release from solid matrices based on swellable/erodible polymeric hydrogels", presented to 19th International Congress of Chemical and Process Engineering CHISA 2010 and the 7th European Congress of Chemical Engineering ECCE-7, 28th August – 1st September 2010, Prague (Czech Republic) - SPEAKER

Dalmoro A.; Galdi I.; Lamberti G.; Titomanlio G.; Barba A.A.; d'Amore M.. "Targeted oral drug delivery by pH-sensitive microparticles", presented to 19th International Congress of Chemical and Process Engineering CHISA 2010 and the 7th European Congress of Chemical Engineering ECCE-7, 28th August – 1st September 2010, Prague (Czech Republic) - SPEAKER

Galdi I.; **Dalmoro A.**; Lamberti G.; Titomanlio G.; Barba A.A.; d'Amore M. "Swelling, erosion and drug release in hydrogel based solid matrices", presented to 7th World Meeting on Pharmaceutics, Biopharmaceutics and Pharmaceutical Technology, 8th-11th March 2010, Valletta (Malta) - SPEAKER

Dalmoro A.; Galdi I.; Lamberti G.; Titomanlio G.; Barba A.A.; d'Amore M. "PH-sensitive microparticles for enteric drug delivery by solvent evaporation from double emulsion", presented to 7th World Meeting on Pharmaceutics, Biopharmaceutics and Pharmaceutical Technology, 8th-11th March 2010, Valletta (Malta) - SPEAKER

Barba A.A., Chirico S., **Dalmoro A.**, Galzerano B., Lamberti G., "Water and drug mass fraction profiles in HPMC/TP matrices", presented to *36th Meeting of Controlled Release Society*, 18th-22nd July 2009, Copenhagen (Denmark) - *SPEAKER*

Barba A.A., D'Amore M., <u>Dalmoro A.</u>, Lamberti G., Titomanlio G., "Enteric coated micro-particles for targeted and controlled release", presented to *36th Meeting of Controlled Release Society*, 18th-22nd July 2009, Copenhagen (Denmark) - *SPEAKER*

Chirico S., **Dalmoro A.**, Lamberti G., Russo G., Titomanlio G., "Radial water up-take in pure HPMC tablet analysis and model prediction", proceedings of Pharmaceutical Sciences World Congress, 22th -25th April 2007, Amsterdam (Netherlands)

Acknowledgments

I would like to thank all the people who helped and supported me during these years of my Ph.D-experience.

A warm thanks to my supervisors for giving me the opportunity to carry out an interesting and pleasant work. In particular, much gratitude to prof. Matteo d'Amore for being a valuable reference point thanks to his knowledge, experience and humanity. A special "Thanks" to prof. Anna Angela Barba, who was not only a coach and a guide, but especially a friend and a confidant, the best I could hope!

Thanks also to prof. Gaetano Lamberti for having taught me love for research and for his continuous presence, only apparently offstage, but ever comforting.

I gratefully acknowledge prof. Giuseppe Titomanlio, especially for our sketch about my final Ph.D. presentation, the pharmacists and "the balance is not closed"!

I want to express my gratitude to my scientific referees, prof. Roland Bodmeier and prof. Nadia Passerini, for their availability to take part of my scientific committee and for their valuable suggestions.

A great thanks to my PhD-mate, Sara, for sharing the good and the bad situations, "the corner of mess-ups", the Gricu experience and the trips all over the Europe!

Thanks also to Valentina: we shared troubles and happiness since bachelor degree up to this important goal.

This Ph.D. has given me the opportunity to meet nice people for funny moments: the Gricu-mates...some of them became "my business partners".

Many thanks go to my two Sisters, Barbara & Monica: the lab T5a brought us together and it made us inseparable.

A kind thanks to the girls of lab. Impianti & Processi, who cheered up the work in the lab, and not only!!! Especially thanks to the "mad group" composed of Amanita, Clara, Enza, Giovanna, Margherita e Michela, headed by Giovanna: in her discovered an Authentic friend! I can't forget the two official "blighters" of the lab, Peppe and Marco, who made lighter work days with our coffees and lunches.

Here I come to the most important people of my life: my parents...thanks to them I have become what I am nowadays, and Gianni, who always supported me in everything, giving me serenity and love.

Finally I thank all the people with whom I have shared experiences, "mess-ups", a coffee, a pizza or a chat.

Simply thanks again to people who have believed and still believe in me...